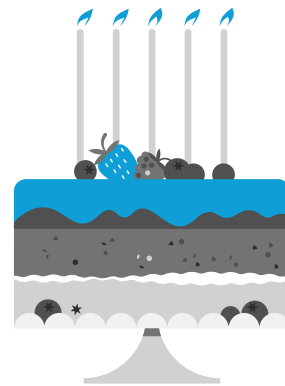
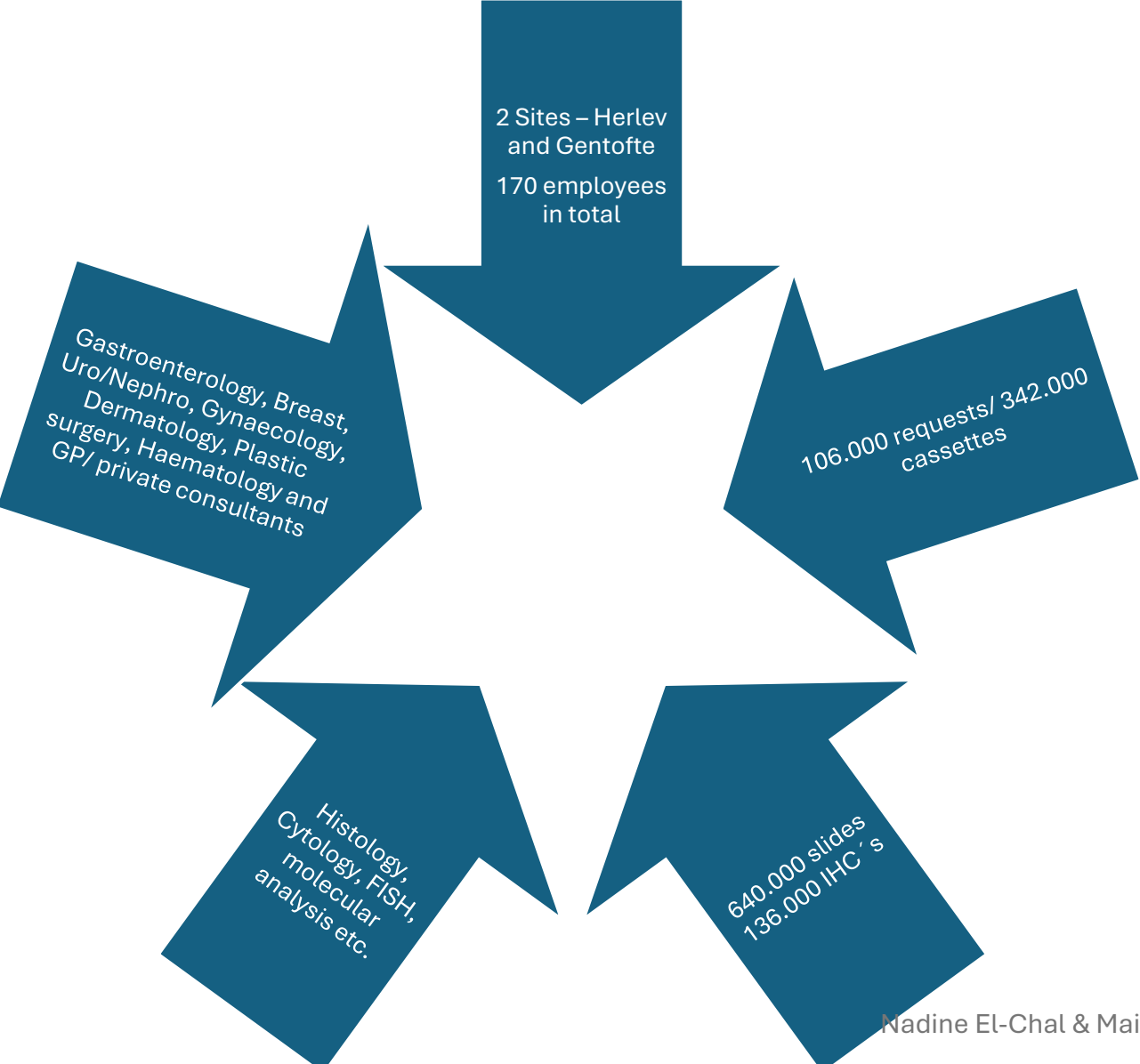


AS410 in the Department of Pathology at Herlev and Gentofte Hospitals

- Almost 10 years with AS410, the beginning, the middle and now...



Our department



Purpose of automation:

- Creating value by sufficient turn around times and high-quality results
- Lack of biomedical lab. scientists
- Work-environment
- Digital pathology

2016 : First in Europe

- **At the time:**
 - Completely new equipment
 - Not installed in routine laboratory anywhere, no prior knowledge to rely on
 - Simple robot, one section per slide online, IHC and SN off-line (lots of manual entering/re-labelling) – (full on-line 2022?)
 - Section time 3 min./block
 - Trimming +/-
 - Only medium/large size nets..
- **Primary goal:**
 - Sectioning all our IHC control slides
 - Sectioning routine, medium and large blocks

IMPLEMENTATION OF AS410 IN 2016

PROJECT GROUP:

2 pathologists & 3 BLS

TESTING:

1. Different net types
2. Tissue processed on Express & VIP
3. Cassettes trimmed on rotation & sledge microtomes
4. Cutting angles 60° and 90°
5. Cutting thickness 3-5µm
6. Frequency of changing blades

EVALUATION:

Sections viewed under microscope according to angle and thickness.

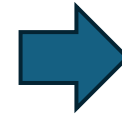
ULTIMATELY

Standardized program ready

TRAINING:

Small group of BLS

TEST PERIOD 2 MONTHS



Thickness 4 µ

Parameter NO.	Cut angle	Drip rate	Knife change	Dripping spot	Mount strength	Mount position
1 STANDARD 9 O M	90°	12µl	Block 18	○●●●●●●●	Medium	45mm
2 STANDARD 6 O M	60°	12µl	Block 12	○●●●●●●●	Medium	45mm
3 STANDARD 9 O L	90°	12µl	Block 18	○●●●●●●●	Low	45mm
4 STANDARD 6 O L	60°	12µl	Block 12	○●●●●●●●	Low	45mm
5 SNMM (Extension 40s)	60°	12µl	Block 1	○●●●●●●●	Medium	45mm
6 NORMAL	60°	12µl		○●●●●●●●	Medium	45mm
7 IHC KONTROL 1	90°	14 µl	Section/1 1 0 0	○●●●●●●●	Medium	22mm
8 IHC KONTROL 2	60°	14µl	Section/1 1 0 0	○●●●●●●●	Medium	22mm
9 Dual slide mounting	60°	First 12µl Second 14µl		○●●●●●●●	Medium	24mm

IHC Control slides – 2 weeks

- How to get enough slides from each block
- How to place tissue in the block
- Mount position

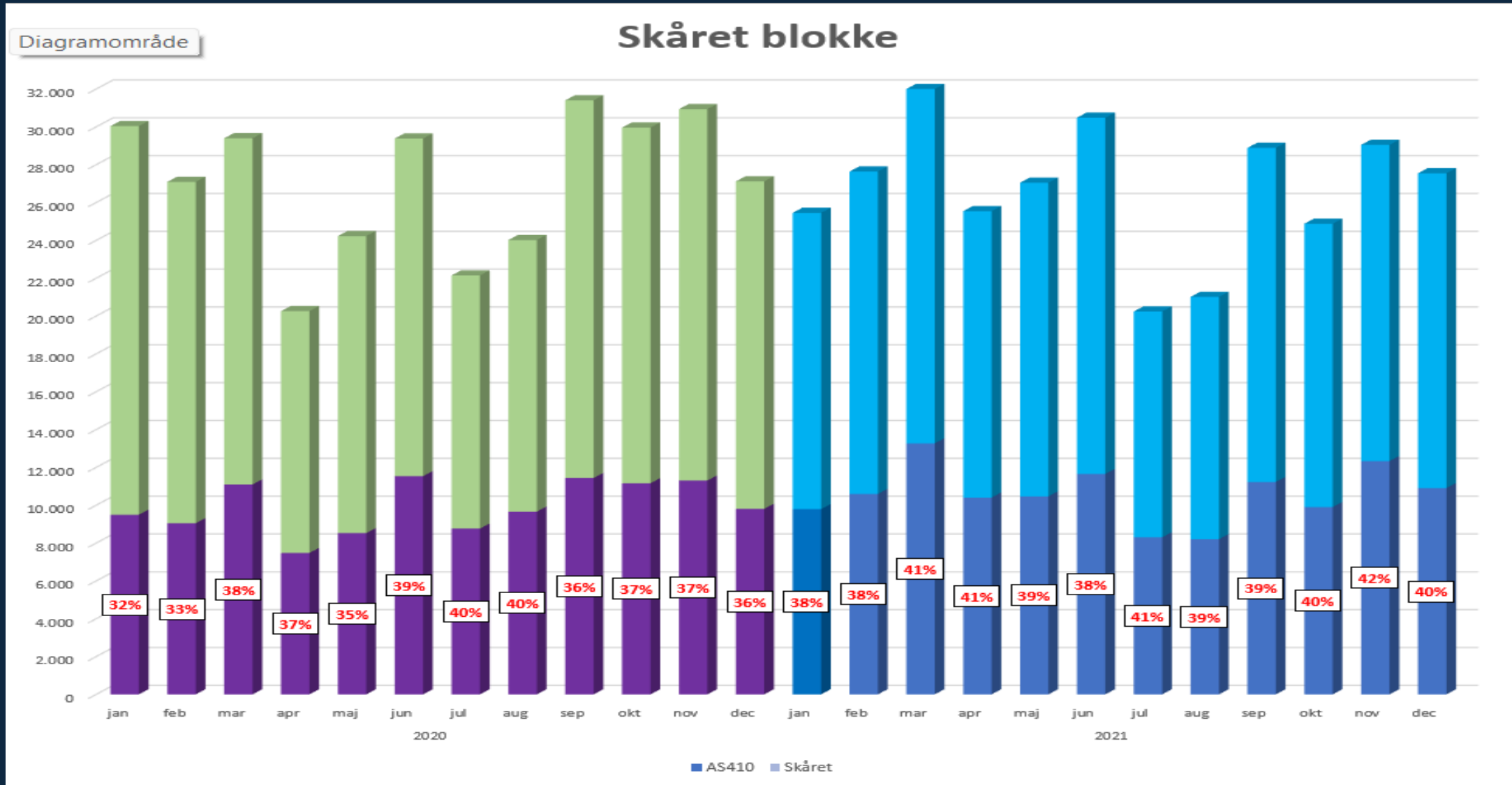


IHC controls (goal: min. 400 slides/block)

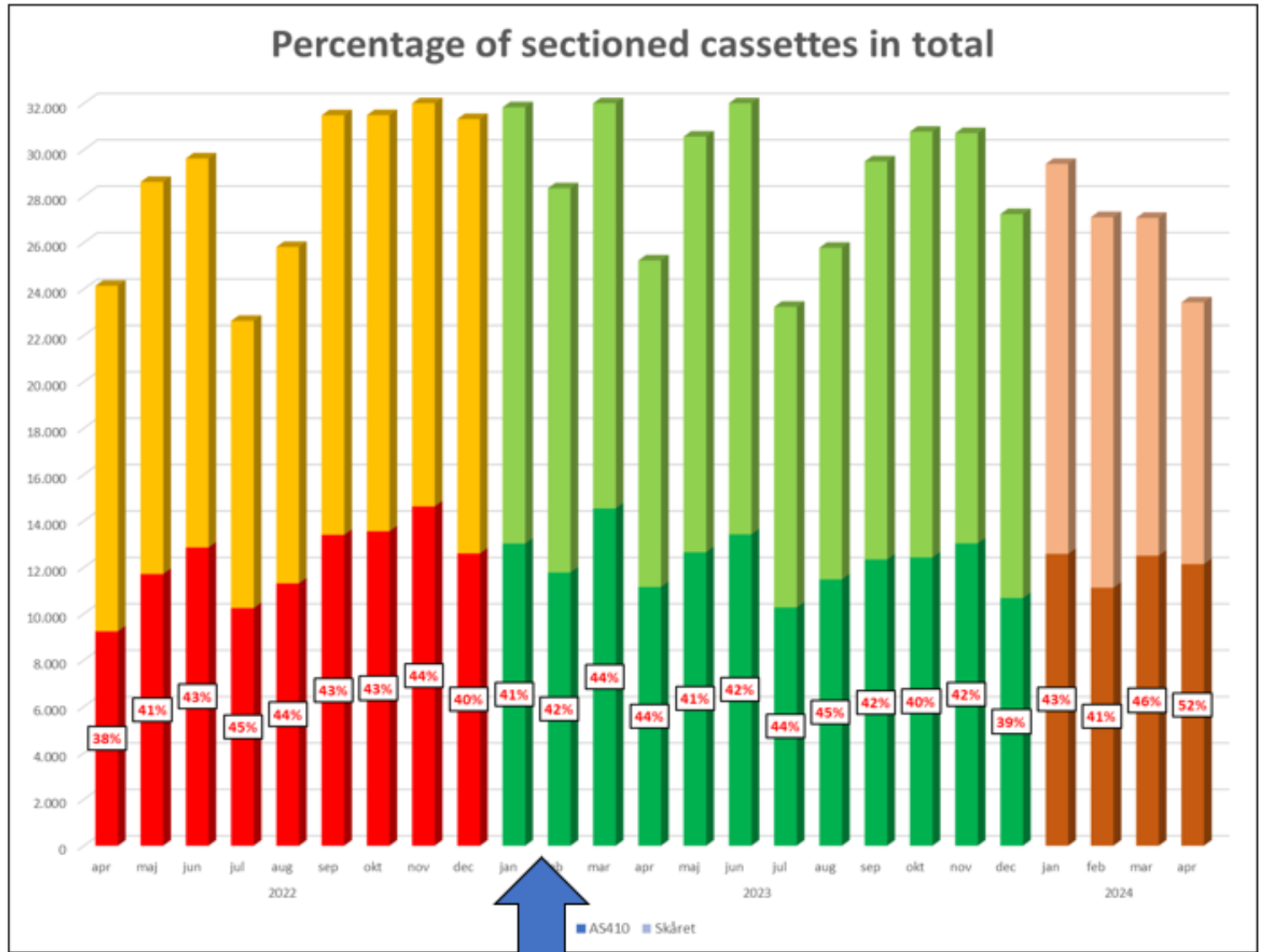
The process

- Collaboration with pathologists crucial
- Training on the AS410
- Learning to read the reports and react appropriately
- Learning how to trim manually
- Implementing new workflow/new function in the lab
- Implementing status changes in order to secure traceability
- Arranging the workspace
- Within 2 weeks we were sectioning IHC controls
- After careful testing/validation we were sectioning medium size nets within 2 months
- Within 6 months we were sectioning approximately 30% of our production on AS410
- A new workflow was implemented!

Results from 2020/2021 – 4th unit installed Q4 2020

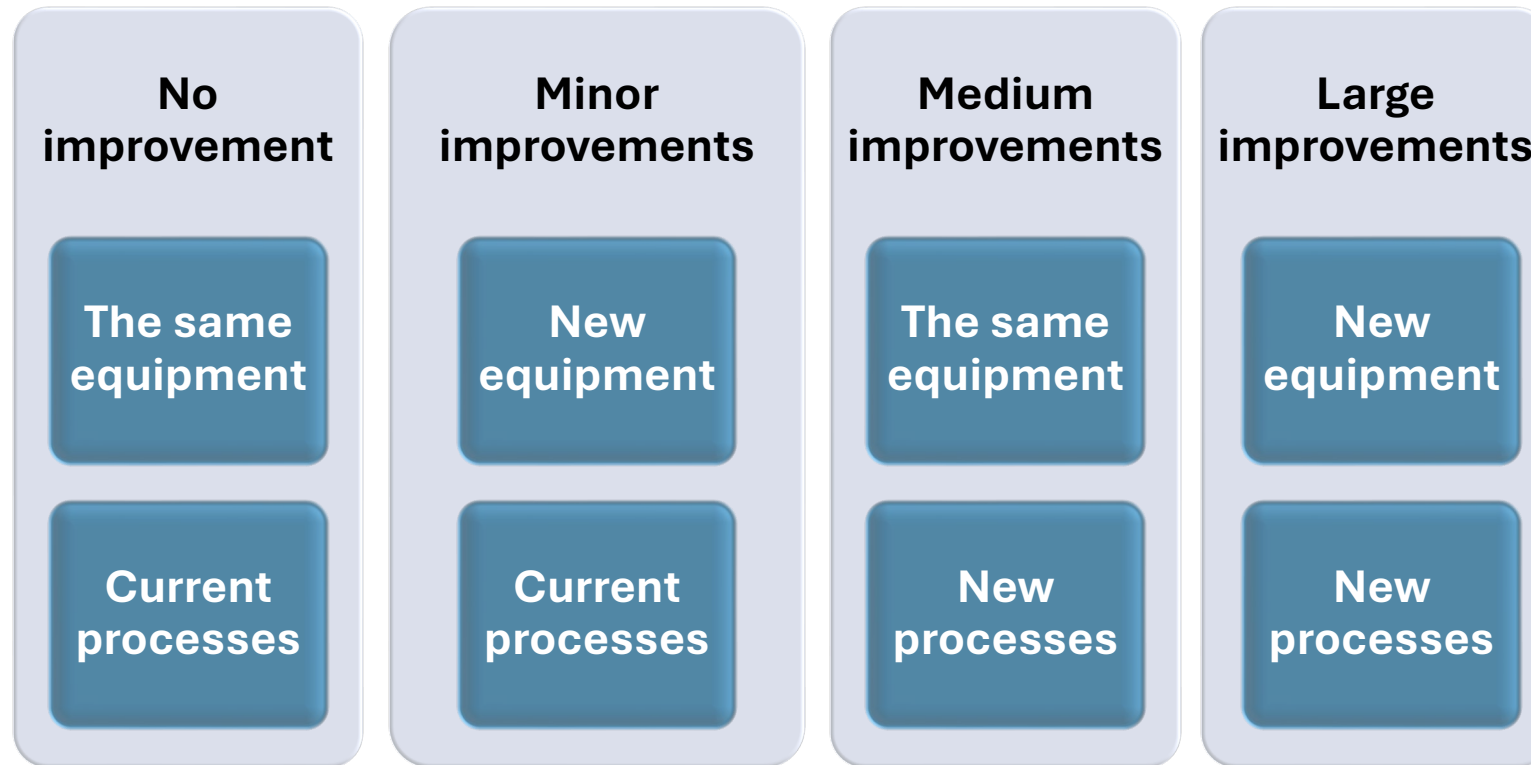


Time passes –
2022 5th unit
installed: Still
no change!



Unit no 5 installed

Our experience when implementing new equipment:



2024: We had to optimize the workflow (full process):

How to optimize?

- LEAN toolbox:
- Value
- Valuestream
- Flow
- Pull
- Perfection
- 5S (incl standardize)

Draw workflow

Optimize sectioning protocols – 5 why and 1 how?

Standardize: Slide flow and organisation of workspace

Standardize grossing, tissue processing, embedding and trimming

Use shortened polish settings

Basically – use LEAN toolbox (or this and following slides;-)

1. Review of workflow

Workflow AS410 – Herlev

	Kl. 7	8	9	10	11	12	13	14	15	16
AS410 - 1	Immun	SNMAM 3 pr. timer							NAT optil 96	96 blokke
		SNMM 2,3 pr. time							Evt. SN ubegrænset	
		SNGYN								
AS410 – 1 tirsdag Hist	Blokke klar	24	48	72	96	120	144	168		168 blokke
	Blokke tilskæres		24	48	72	96	120	144		144 blokke
AS410 - 2	Blokke klar	24	48	72	96	120	144	168	NAT optil 96	264 blokke
	Blokke tilskæres		24	48	72	96	120	144		240 blokke
AS410 - 3	Blokke klar	24	48	72	96	120	144	168	NAT optil 96	264 blokke
	Blokke tilskæres		24	48	72	96	120	144		240 blokke
AS410 - 4	Immun	SNMAM 3 pr. timer							Kontroller 400 glas	
		SNMM 2,3 pr. time								
		SNGYN								
AS410 - 5	Blokke klar	24	48	72	96	120	144	168	NAT optil 96	264 blokke
	Blokke tilskæres		24	48	72	96	120	144		240 blokke
									1056/888/816 hist blokke + SN optil 18 pr. AS pr. dag + nat	

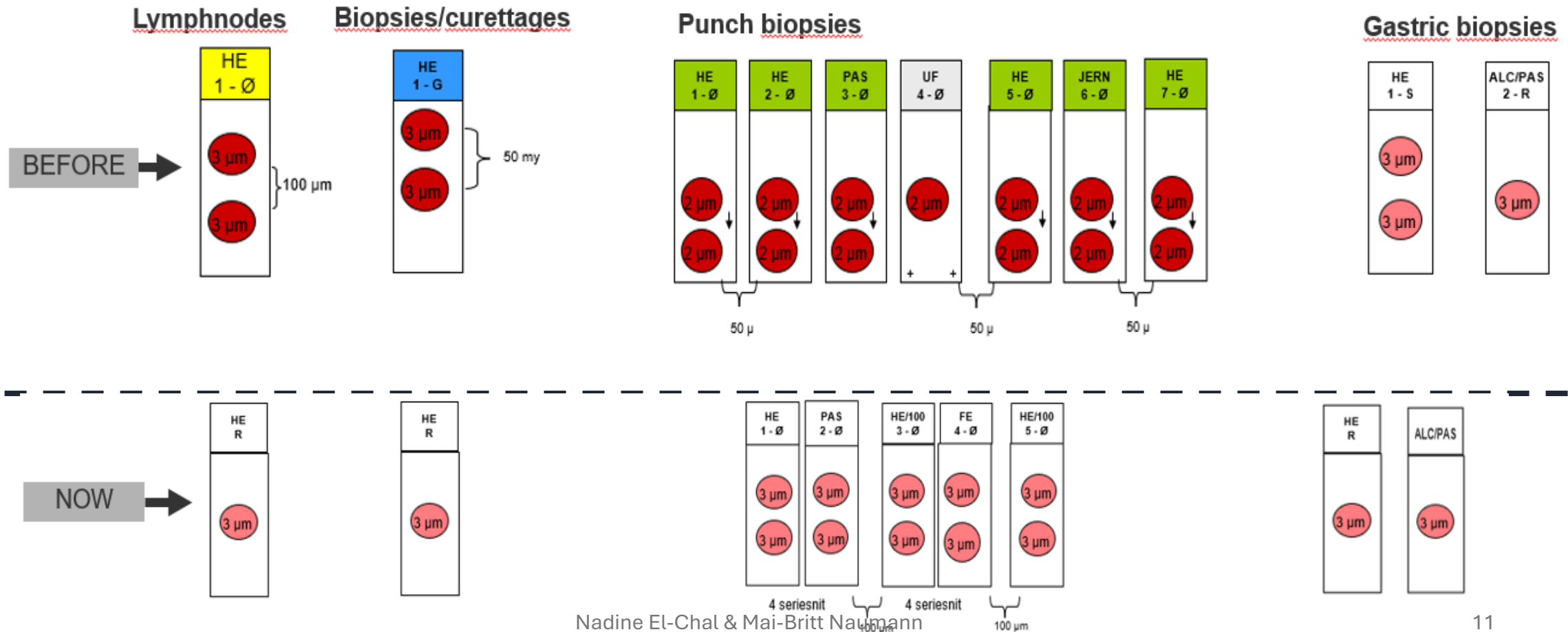
Hist



Immun

2. Review of protocols

- Meetings with pathologists
- Introduction to new protocols
- Workshops

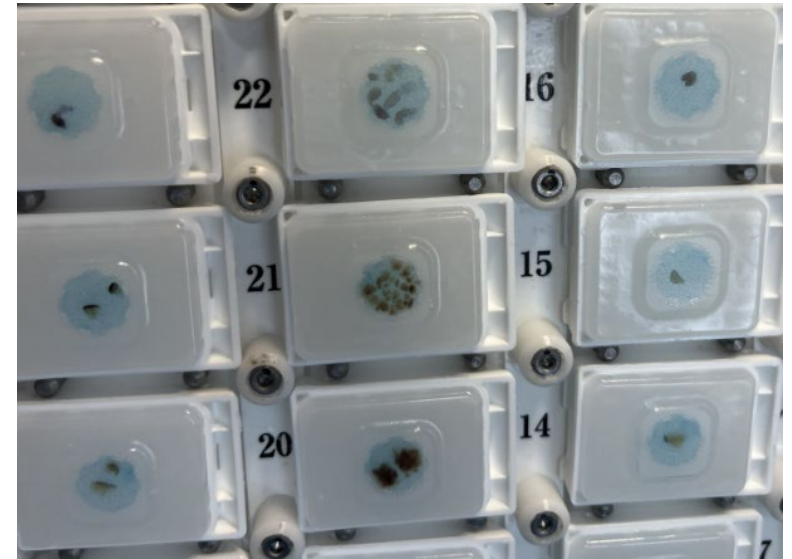


Organ specialities

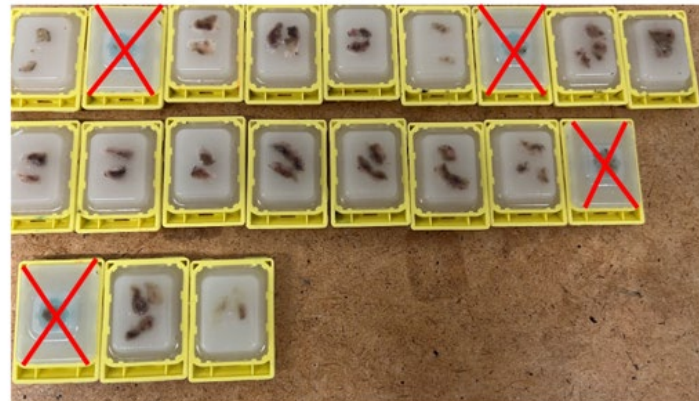
- Breast
- Gastroenterology
- Urology/nephrology
- Gynaecology
- Dermatology/ plastic surgery
- Samples from private clinics

In 2016: 'preferable to use only medium size nets', and small nets were not sectioned on AS until 2024

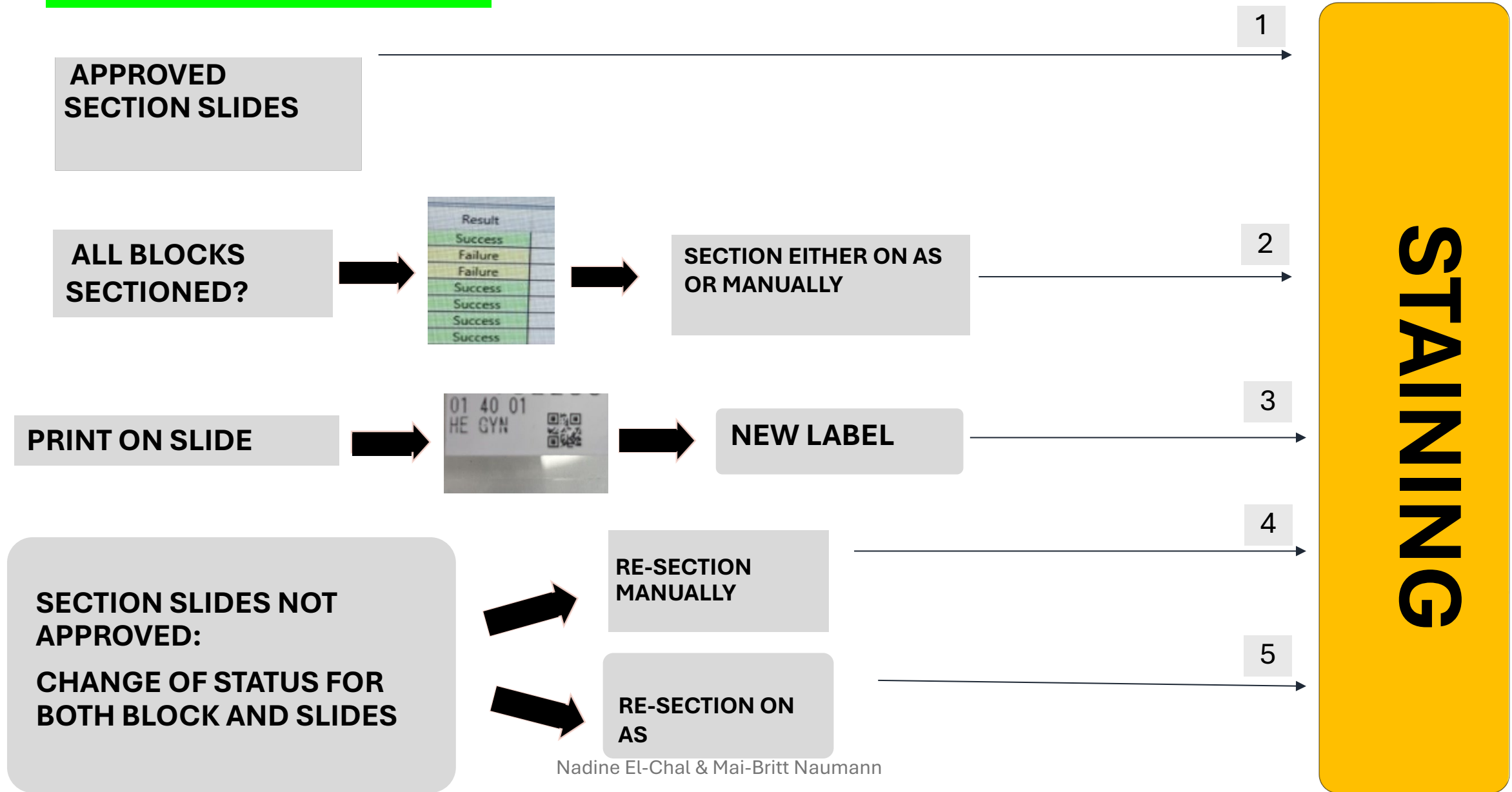
Small size: Oct. 2024



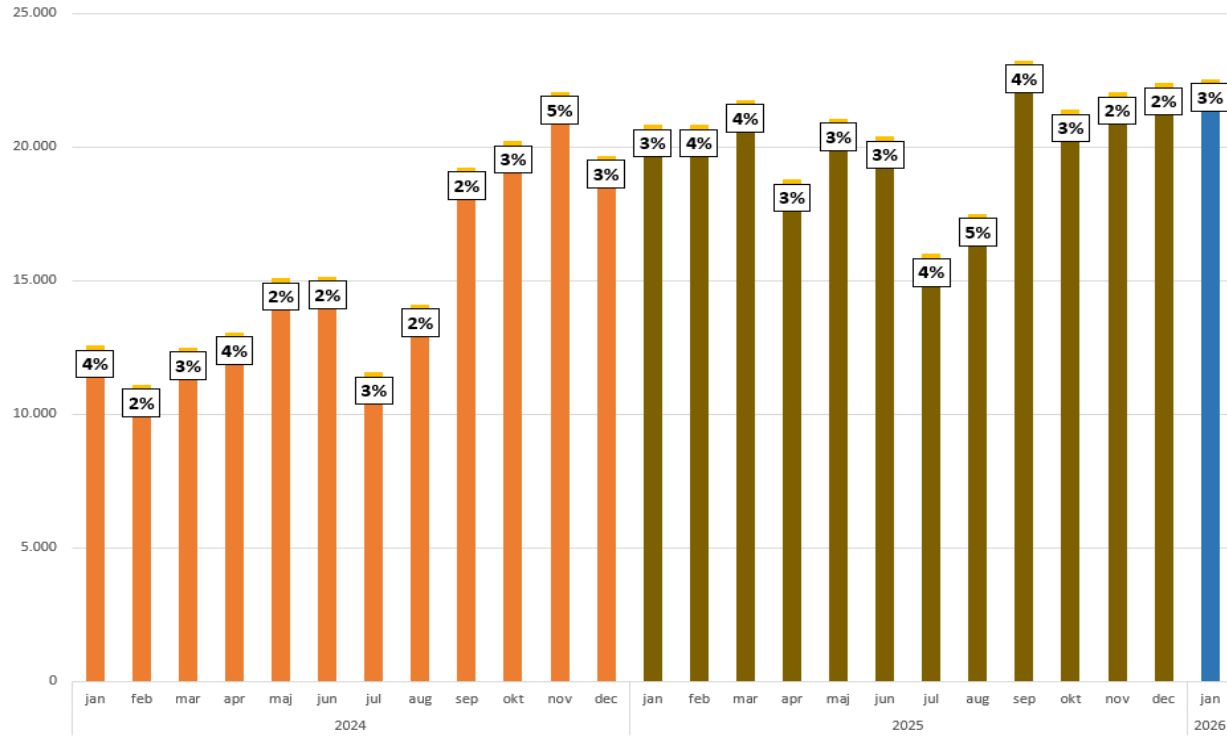
Blocks from an Intestine sectioning of 1 patient



3. Review of slide flow



4. Re-cuts?

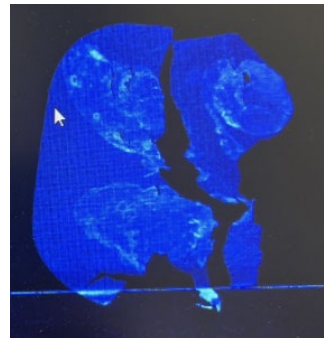
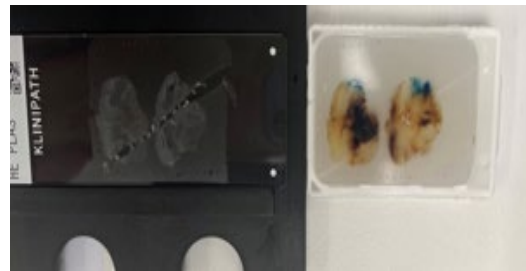


Re-cuts 3-6%

2023	dec	12.594	11.883	711	6%
	jan	13.022	12.579	443	3%
	feb	11.776	11.220	556	5%
	mar	14.535	13.892	643	4%
	apr	11.147	10.609	538	5%
	maj	12.650	12.034	616	5%
	jun	13.420	12.946	474	4%
	jul	10.269	9.826	443	4%
	aug	11.476	11.153	323	3%
	sep	12.337	11.899	438	4%
	okt	12.425	12.025	400	3%
	nov	13.031	12.613	418	3%
dec	10.670	10.295	375	4%	

Re-cuts 2-5%

2025	jan	20.833	20.131	702	3%
	feb	20.833	20.014	819	4%
	mar	21.759	20.995	764	4%
	apr	18.776	18.238	538	3%
	maj	21.061	20.411	650	3%
	jun	20.398	19.856	542	3%
	jul	15.991	15.399	592	4%
	aug	17.495	16.685	810	5%
	sep	23.237	22.366	871	4%
	okt	21.393	20.766	627	3%
	nov	22.053	21.506	547	2%
	dec	22.382	21.948	434	2%



? Good/no good ?

Madine El-Chal & Mai-Britt Naumann

5. Focus on grossing

- Tissue (Thickness & replacement of tissue)
- The 'right' net
- Closing the net correct
- Use of small nets



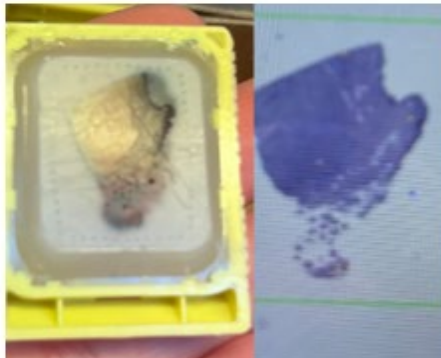
6. Embedding

- Levelling (one plane)
- No bubble



7. Pre-trimming

- All tissue represented



8. Polish setting adjustment (September 2025)

Polish units have impact on time, polish steps shorten

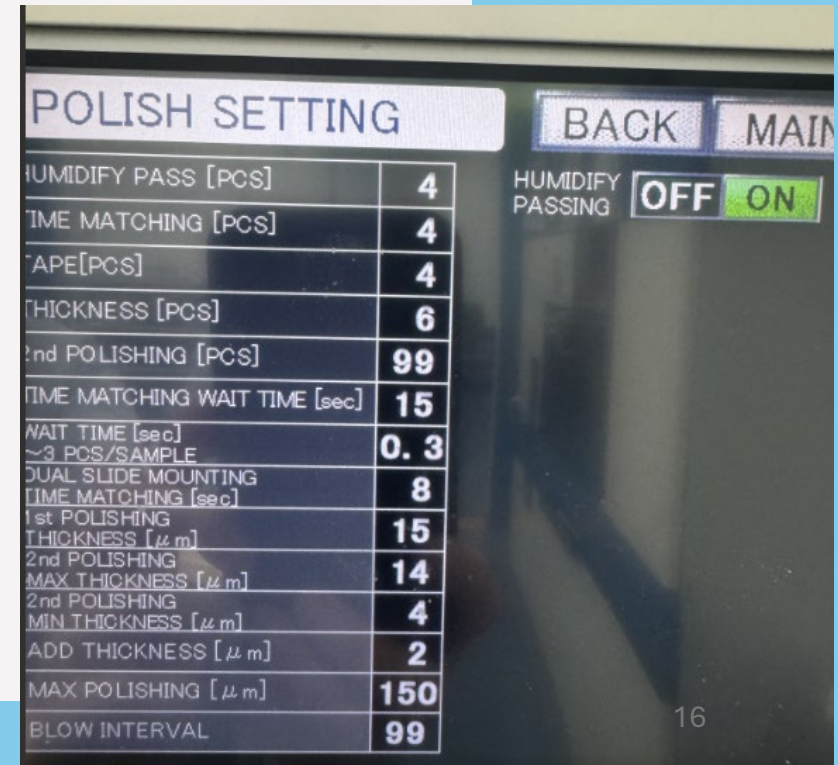
PolishCut No.01 Thickness = 5[μm] (2nd Polishing)
 PolishCut No.02 Thickness = 5[μm] (2nd Polishing)
 PolishCut No.03 Thickness = 5[μm] (2nd Polishing)
 PolishCut No.04 Thickness = 5[μm] (2nd Polishing)
 PolishCut No.05 Thickness = 5[μm] (2nd Polishing)
 PolishCut No.06 Thickness = 5[μm] (2nd Polishing)
 PolishCut No.07 Thickness = 5[μm] (2nd Polishing)
 PolishCut No.08 Thickness = 5[μm] (2nd Polishing)
 PolishCut No.09 Thickness = 5[μm] (2nd Polishing)
 PolishCut No.10 Thickness = 5[μm] (2nd Polishing)
 PolishCut No.11 Thickness = 5[μm] (2nd Polishing)
 PolishCut No.12 Thickness = 5[μm] (2nd Polishing)
 PolishCut No.13 Thickness = 4[μm] (2nd Polishing)
 PolishCut No.14 Thickness = 6[μm] (MakingThickness+Additional Polishing) <Tape Mount>
 PolishCut No.15 Thickness = 4[μm] (MakingThickness Polishing) <Tape Mount>
 PolishCut No.16 Thickness = 4[μm] (MakingThickness Polishing) <Tape Mount>
 PolishCut No.17 Thickness = 4[μm] (MakingThickness Polishing) <Tape Mount>

2nd polish max from 5μm → 7μm
 Saves 4 polish steps

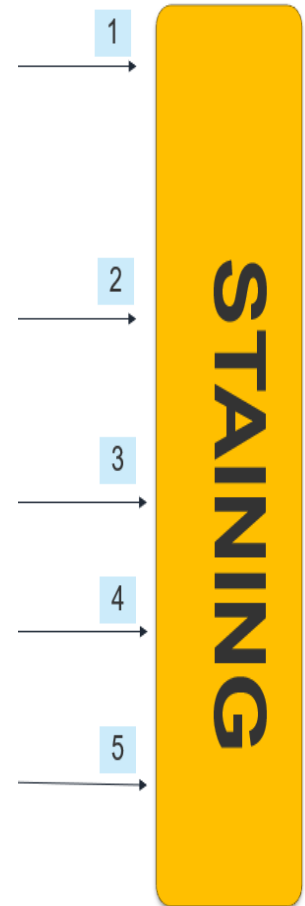
PolishCut No.01 Thickness = 7[μm] (2nd Polishing)
 PolishCut No.02 Thickness = 7[μm] (2nd Polishing)
 PolishCut No.03 Thickness = 7[μm] (2nd Polishing)
 PolishCut No.04 Thickness = 7[μm] (2nd Polishing)
 PolishCut No.05 Thickness = 7[μm] (2nd Polishing)
 PolishCut No.06 Thickness = 7[μm] (2nd Polishing)
 PolishCut No.07 Thickness = 7[μm] (2nd Polishing)
 PolishCut No.08 Thickness = 7[μm] (2nd Polishing)
 PolishCut No.09 Thickness = 7[μm] (2nd Polishing)
 PolishCut No.10 Thickness = 6[μm] (MakingThickness+Additional Polishing) <Tape Mount>
 PolishCut No.11 Thickness = 4[μm] (MakingThickness Polishing) <Tape Mount>
 PolishCut No.12 Thickness = 4[μm] (MakingThickness Polishing) <Tape Mount>
 PolishCut No.13 Thickness = 4[μm] (MakingThickness Polishing) <Tape Mount>

2nd polish max from 5μm → 10μm
 Saves 6 polish steps

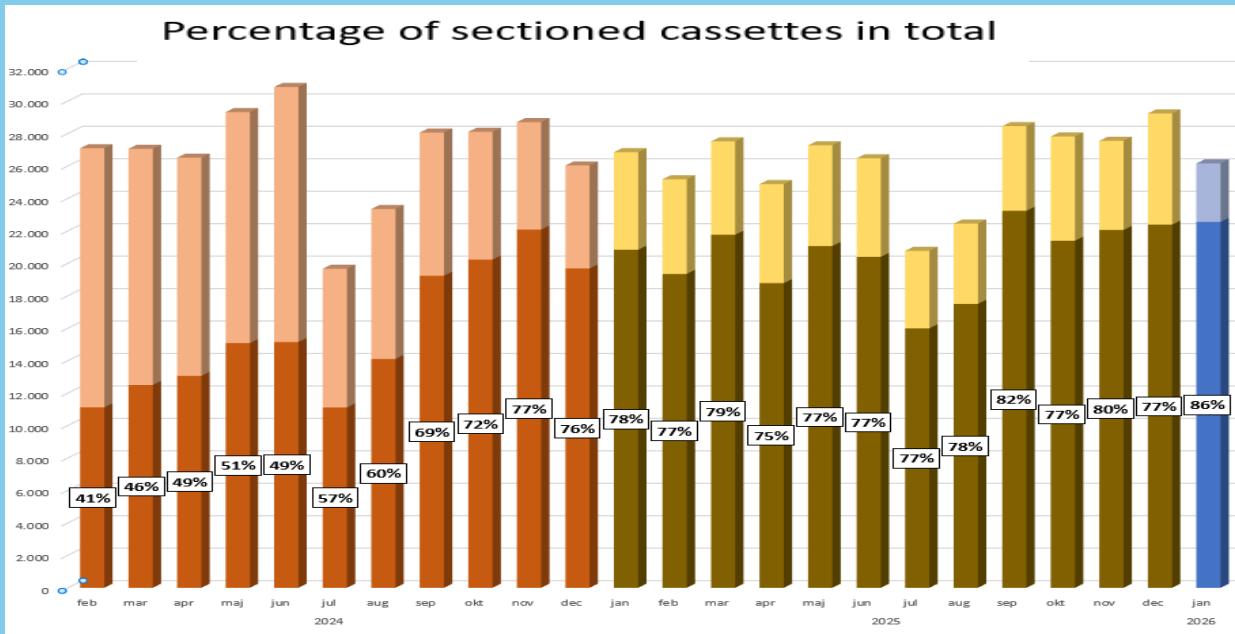
PolishCut No.01 Thickness = 10[μm] (2nd Polishing)
 PolishCut No.02 Thickness = 10[μm] (2nd Polishing)
 PolishCut No.03 Thickness = 10[μm] (2nd Polishing)
 PolishCut No.04 Thickness = 10[μm] (2nd Polishing)
 PolishCut No.05 Thickness = 10[μm] (2nd Polishing)
 PolishCut No.06 Thickness = 10[μm] (2nd Polishing)
 PolishCut No.07 Thickness = 4[μm] (2nd Polishing)
 PolishCut No.08 Thickness = 6[μm] (MakingThickness+Additional Polishing) <Tape Mount>
 PolishCut No.09 Thickness = 4[μm] (MakingThickness Polishing) <Tape Mount>
 PolishCut No.10 Thickness = 4[μm] (MakingThickness Polishing) <Tape Mount>
 PolishCut No.11 Thickness = 4[μm] (MakingThickness Polishing) <Tape Mount>



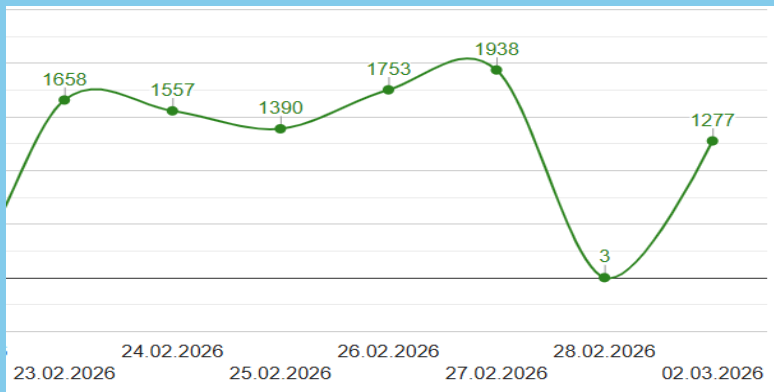
9. Organization in lab



2026:



Blocks sectioned /day



5 units AS-410M + AT-192



lab. Scientists



Maintenance	15 min/unit
Morning unload + QC	1,5 hour
Overall time	
Processing hours each unit	Day 7-9½ (+4 hour nightrun)

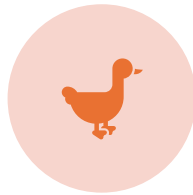


Blocks sectioned/day- 80-87% on AS410

What I would do different:



Careful yet brave



Careful validation can trigger overall carefulness and hinder improvement



Deep dive into sectioning protocols – old habits die hard



Pay more attention to workplace culture



Share knowledge, ask questions ...reach out (we are many labs on this journey)



Collect data for validation reports



Few specialists on AS410 – a larger group handling on/off procedure



Teach every tech how to plan their workday with AS410

Future goals-almost there:

- 80-90% of all cassettes on AS410 ✓
- Interval-Routine ✓
- Punch biopsies on AS ✓
- Faster section time ✓
- Mixed running programs/run
- Kaizen 😊

Thank You!

- Feel free to reach out:

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Mai-Britt.Naumann@regionh.dk

